

Review Article

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Biofertilizers and Biopesticides: Microbes for Sustainable Agriculture diseases management of crops: a review Preeti Vashisht¹, Prahlad¹ and N.K. Yadav²



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ABSTRACT

Agriculture development initiatives on the part of humans have long been documented. Since learning about the significance of this area, farmers and researchers have not stopped looking for strategies and solutions to increase agricultural output and quality while also shielding it from potential threats and stress. In place of agrochemicals, mechanisms using microorganisms as biofertilizers, and biocontrol agents have recently gained popularity. Utilizing advantageous microorganisms is an environmentally benign tactic that plays a significant part in promoting plant growth and in the biocontrol of plant diseases. Reduced chemical inputs and the usage of harmful pesticides in agricultural soils may be possible with a greater understanding of how these bacterial communities are used. The focus of the current review is on plant growth-promoting bacteria (PGPB), and it provides a summary of their function in soil fertilization and plant protection with a focus on their methods of action. This chapter includes a number of PGPB examples that were taken from the literature. Examples of how these bacteria have been used in agriculture are also included in this review.

Keywords: Agriculture, Biofertilizers, Biopesticides, Microorganisms, Soil, biocontrol, fertilization, chemical

1. Introduction

Human efforts to boost food production and cut expenses have detrimental repercussions on a variety of levels. Chemical solutions are seen as the most practical solution, but their widespread usage is harmful to both human health and the environment (1,2). Other obstacles to agriculture besides chemical products include salinity, heavy metals and desert regions. In reality, agriculture has intensified because of market globalization, rapid population growth and rising living standards in the most industrialized nations. This intensification is based on mechanization and sophisticated agronomic techniques, which have increased crop output. Due to environmental harm, intensive agriculture is coming under increasing criticism. These environmental degradations have led to a number of new agricultural science problems, including those related to depollution, waste management, rural development, and biological control (3). Plant infections have disastrous consequences on plant health and crop yields, which further endangers food production and ecological stability. Chemical products have recently been the most popular crop protection method among farmers. These substances, often known as biostimulants, frequently provide cutting-edge fertilization and crop protection techniques. The majority of these biologically generated stimulants are created by plant growth-promoting bacteria (PGPB) (4). This collection of microorganisms greatly increases sustainable agriculture by regulating the rhizosphere's biological activity and

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DOI: https://doi.org/10.58321/AATCCReview.2023.11.04.398 © 2023 by the authors. The license of AATCC Review. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/). safeguarding and promoting plant growth and health (5). These substances, often known as biostimulants, frequently provide cutting-edge fertilisation and crop protection techniques. The majority of these biologically generated stimulants are created by plant growth-promoting bacteria (PGPB). This collection of microorganisms greatly increases sustainable agriculture by regulating the rhizosphere's biological activity and safeguarding and promoting plant growth and health. The selection of beneficial bacteria and their byproducts can enhance plant rhizosphere colonization, create a wide range of agricultural prospects, and preserve the environment. In fact, a better knowledge of how these bacterial communities function might enable the usage of toxic pesticides and chemicals in agricultural soils to be reduced (6).

2. Plant Growth-Promoting Bacteria

The microorganisms, which are mainly bacteria from the Arthrobacter, Azotobacter, Azospirillum, Bacillus, Enterobacter, Pseudomonas, Serratia, and Streptomyces spp. genera, encourage the growth of plants. Moreover, these bacteria protect plants from biotic and abiotic threats (7,8,9,10). These microbes can grow in number and compete with other soil microbes for nutrients and rhizospheric space. Rhizobacteria boosting plant nodulation and plant growth-promoting bacteria (PGPB) are the names given to these microorganisms (11,12). Plant growth-promoting rhizobacteria, or PGPRs, are the subset of PGPBs that have received the greatest research attention (13). Yet, unlike other rhizosphere bacteria, these bacteria provide plants with a number of beneficial effects through a variety of direct and indirect mechanisms. These bacteria infiltrate the plant rhizosphere by using root exudates as nutrient substrates (14,15,16). Auxins, cytokinins, gibberellins, nitrogen fixation, phytohormone synthesis (auxins, cytokinins, and gibberellins), siderophores production, and inhibition of ethylene synthesis by the enzyme 1-aminocyclopropane-1-carboxylic deaminase

(ACC-deaminase) are examples of direct mechanisms (17, 18, 19). Some examples of indirect mechanisms include the production of hydrolytic enzymes, the inhibition of pathogenproduced enzymes or toxins, the induction of plant resistance mechanisms, and the suppression of phytopathogenic agents through competition for nutrients and space (19, 20, 21). PGPB diversity varies widely depending on the kind of plant, the kind of soil, and the availability of nutrients. Of the PGPBs that have been discovered, Pseudomonas and Bacillus are the most widespread and thoroughly studied. Strains from the genera Aeromonas, Azospirillum, Azotobacter, Arthrobacter, Clostridium, Enterobacter, Gluconacetobacter, Klebsiella and Serratia are also included in the PGPB group (22,23,24,25). In addition to being efficient biofertilizers and having the enzymes (lipase, esterase, protease, phosphatase, urease, chitinase, and amylase) to hydrolyze all varieties of organic polymers, PGPB has the capacity to enhance the growth of all plant growth parameters (seed germination, root and shoot length and weight, leaf area, and chlorophyll content) (26,27). As a result, they facilitate nutrient uptake by plants and help to enrich the soil with nutrients (23,28). Rhizoremediers (degrade organic pollutants and lessen their toxicity), phytostimulators (stimulate the growth and the development of different plant growth parameters), phytopesticides (protect the plants from various aggressions caused by phytopathogenic agents), and biopesticides (degrade organic pollutants and mitigate their toxicity) are some applications for PGPB (29,10,27).

2.1 PGPB as Biofertilizers

Biofertilizers are substances that contain useful microorganisms (living or dormant) that have the ability to enrich soil with nutrients and stimulate plant growth when applied to soil. They do this by improving nutrient uptake, nutrient efficiency, tolerance to abiotic stress, and crop productivity and quality. Biofertilizers are preparations of active or dormant cells of strains with the ability to fix nitrogen, solubilize phosphate, and produce cellulase (30). To improve soil and boost plant productivity, PGPBs employ a number of techniques, such as nitrogen fixation, phosphate solubilization, siderophores, and hydrolytic enzyme production (31,32,33). In particular, nitrogen is produced by bacteria that fix nitrogen (34), iron is produced by bacteria that produce siderophores, sulfur is produced by bacteria that oxidise sulfur (35), and phosphorus is produced by bacteria that solubilize phosphate. Plants can more readily absorb nutrients thanks to biofertilizers (36). We may utilize these beneficial microorganisms and boost crop productivity by comprehending how these bacteria work with other rhizospheric germs (37).

2.1.1 Enzymatic Activities

The fact that microbial enzymes are used so frequently in biotechnology—including agro-food, detergents, textile, pharmacy, and molecular biology—is due to the fact that these microorganisms have attracted so much attention in agriculture as biofertilizers. Enzymes including proteases, esterases, lipases, amylases, and cellulases are crucial to the agricultural business because they play a crucial part in soil fertilization through the breakdown of organic polymers (38). During these processes, these microbes release nutrients such as phosphate, iron, carbon, nitrogen, potassium, and sulphur into the soil, which are then utilized by plants and other soil microorganisms Cell wall-degrading enzymes, such as those secreted by the PGPR, such as glucanases, chitinases, cellulases, and proteases, are one of the main strategies used by biocontrol agents to control plant diseases (39,40). The essential conversion of cellulose into carbon dioxide or methane by bacteria is the primary role of cellulose degradation in the carbon cycle (41). There is a lot of interest in the prospect of converting agricultural waste utilizing cellulases made by soil microorganisms. The number of nodules increases as a result of this enzyme making it simpler for these bacteria to access the intercellular space or the root hair (42). Bacteria including Pseudomonas aeruginosa, Bacillus prodigiosus, Bacillus pyocyaneus, and Bacillus prodigiosus were found to have lipases and esterases in 1901. Protein breakdown by microbial proteases has a significant impact on the nitrogen cycle in soil because it makes nitrogen available to plants and microorganisms (43). It is well known that the creation of lytic enzymes like proteases is one of the indirect strategies employed by PGPR to eliminate undesirable bacteria (44). Proteases are utilized in the disintegration of worm cuticles (45). The nitrogen cycle in soil depends on microbial proteases because it makes nitrogen available to plants and microbes (43). It is well known that one of the indirect methods employed by PGPR to eliminate harmful microorganisms is the development of lytic enzymes such as proteases (44). Several authors have shown the effect of microbial proteases on biological control (39,40). Pythium ultimum, a phytopathogenic fungus, is suppressed in the sugarcane rhizosphere by extracellular protease produced by *Stenotrophomonas maltophilia* W81,(46). The protease activity may indirectly alter the synthesis of auxin by releasing amino acids like tryptophan, a precursor for the synthesis of the IAA (47). A sizable amount of urea is continuously released into the environment as a result of biological processes. Urease is an extracellular enzyme that hydrolyzes urea into CO₂ and ammonia, which accounts for 63% of all soil activity (NH3). Due to the fact that the rate of organic matter synthesis has an impact on its concentration, it is employed as a soil quality indicator (48). Since Rotini, first reported on the activity of urease in soils, it has attracted a lot of attention (49). According to Polacco, microbes and plants that produce both intracellular and extracellular enzymes are the primary producers of soil urease (50).

2.1.2 Nitrogen Fixation

Nitrogen is one of the most important factors in plant growth. As the majority of this element exists as gas (N2), neither plants nor animals can utilize it (51). In order to meet the plants' needs for nitrogen and to improve productivity, agriculture has become dependent on artificial fertilizers; however, the harm these products caused was substantially more than their beneficial effects. A study on the harm that synthetic nitrogen fertilizers can cause to human health as well as the environment was released by the US National Institutes of Health. It was postulated that increased nitrate levels in drinking water may contribute to cancer and that cardiac disorders may be becoming more common due to nitrogen-related air pollution (52,53). Farmers consequently depended more and more on biological mechanisms to provide nitrogen to plants. Bacteria repair this element and also change it into ammonia (assimilable form). Examples of free-living bacteria include Azotobacter, Bacillus, Acetobacter, Clostridium, Klebsiella, Corynebacterium, Arthrobacter, Diazotrophicus and Pseudomonas. An illustration of a symbiotic bacteria is Rhizobium (54,55). The benefit of diazotrophic bacteria is that they can exchange the carbon they create in the form of root

exudates for plants' need for nitrogen. Yet, the presence of carbon as an energy source is required for the substantial nitrogen fixation. This calls for the presence of these diazotrophs around plants, either as endophytes, in the rhizosphere, or in the rhizoplane. Nitrogen fixation has the potential to boost soil fertility and productivity. Many studies have examined the inoculation of plants with nitrogen-fixing bacteria, and several authors have demonstrated the role of PGPR in enhanced nodulation in root plants as well as nitrogen fixation (56). By utilizing these microorganisms, the need for chemical fertilizers can be greatly reduced. Azotobacter sp., Paenibacillus polymyxa and the endophytic bacteria Azoarcus sp., Burkholderia sp., Gluconacetobacter diazotrophicus, and Herbaspirillum sp. have all been shown to fix nitrogen, and this ability is closely correlated with their ability to promote plant growth (57). Three nitrogen-fixing bacteria, known as Azotobacter diazotrophicus, Herbaspirillum seropedicae, and Azoarcus spp., are found in the rhizosphere of plants, namely in their roots. Barley, wheat, rice, and sugar cane production are all increased by these bacteria (58). Inoculating plants with nitrogen-fixing bacteria including Azospirillum, Enterobacter, and Rhizobium while they were under axenic conditions increased potato output (59). For instance, Azospirillum sp. TN10 increased the fresh and dry weight of the potato in comparison to control (non-inoculated) plants. It was also discovered that the nitrogen content of the stems and roots of the infected plants had greatly risen.

2.1.3 Phosphate Solubilization

Like nitrogen, phosphorus (P) is an essential nutrient for the growth of plants and a nutrient that restricts plant growth (57). It is an essential component of DNA and adenosine triphosphate (ATP) (60). This element is immobilized in the soil by chemical precipitation, which renders it less soluble and unavailable to plants (61). The majority of the phosphorus taken during the developmental phases is transferred to the fruits and seeds; however, phosphorus-deficient plants show growth retardation (reduction of growth of cells and leaves, disruption of respiration and photosynthesis). In agricultural soils, the solubilization of inorganic phosphates is directly influenced by the activity of microorganisms (62,60). Bacterial species such as Bacillus, Pseudomonas, Rhizobium, Aspergillus and Penicillium can transform phosphorus into a form that plants can utilise (63,64,65). The two main procedures employed by PGPRs to convert phosphate into an assimilable state are solubilization and mineralization. The solubilization is brought on by the release of low molecular weight organic acids like gluconic acid and citric acid (66,67). These molecules lower the pH by chelating the cations attached to the insoluble phosphates and converting them to soluble forms (68). The mineralization process carried out by the release of extracellular enzymes including phosphatase and phytases that catalyze the breakdown of the phosphoric esters. Both of these tactics can coexist in the same bacterial species (66). It has been shown by many writers that bacteria can solubilize phosphate (64). According to Gull et al., there is a significant potential for phosphate solubilization in soil by species of the genera Bacillus and Pseudomonas. Bacteria could be used in crop fields because of their capacity to solubilize phosphate. The species Achromobacter xylosoxidans, Bacillus polymyxa, Pseudomonas putida, Acetobacter diazotrophicus, Agrobacterium radiobacter, Bradyrhizobium mediterraneum, Pseudomonas and other phosphate-solubilizing bacteria can increase the amount of

phosphate in the soil. Pseudomonas species boost plant growth by enhancing their ability to absorb different minerals from the soil and as plant growth promoters. Pseudomonas and other phosphate-solubilizing bacteria can increase the amount of phosphate in the soil (69). Pseudomonas species, increase plant growth by increasing their capacity to absorb various minerals from the soil. Pseudomonas species such as P. cissicola, P. fluorescens, P. pinophillum, P. putida, P. syringae, P. aeruginosa, P. putrefaciens and P. stutzeri that have been identified from the rhizosphere of cereals such as chickpea, corn, and soybean greatly solubilize phosphates. According to several studies, Pisum sativum L. grown in soil deficient in soluble phosphate produces significant quantities of gluconic acid as a result of being stimulated to develop (67). The inoculation of Chickpea by single and dual phosphate-solubilizing Bacillus megaterium (M3) and nitrogen-fixing Bacillus subtilis (OSU-142) has improved all its parameters growth compared to control equal to or higher than N, P, and NP treatments (70).

2.2 PGPB as Biocontrol Agents

Biocontrol, also referred to as biological control or biocontrol, is the employment of living animals to defend plants against various threats. It refers to the practise of controlling some pathogenic insects in entomology with the use of predatory insects or entomopathogenic nematodes. In phytopathology, this statement describes the use of microorganisms to manage hazardous plants and avoid disease. The International Organization for Biological Control (IOBC) defines biological control as the use of live organisms to prevent or reduce harm caused by pests. These microorganisms go by the moniker "Biological Control Agent" (71). Many studies examining the use of these bacteria as pesticide substitutes have shown how crucial a part these biocontrol agents may play in raising horticulture and agriculture productivity (72). Many varieties of bacteria and fungi, especially those belonging to the genera Bacillus, Pseudomonas and Burkholderia, have been referred to as unfriendly microorganisms (73). There is a broad search for novel biological control strategies to stop the formation of phytopathogenic bacteria because of environmental concerns. It has been proposed that the inhibition of phytopathogenic fungi by bacteria may be caused by the synthesis of antibiotics, the secretion of hydrolytic enzymes, the creation of plant resistance, the competition for resources and space, or a combination of these processes (74,75,76,77,78). Changes in environmental parameters (such as pH, plant area, etc.) are one of the mechanisms used by several biocontrol agents to indirectly control plant diseases (79).

2.2.1 Siderophores Production

A protein called a siderophore has the ability to solubilize and bind iron from the soil so that it can be given to plant cells. These low-molecular-weight iron-binding molecules have an affinity for Fe³⁺. This chemical is utilized by PGPRs in the biocontrol of phytopathogens as well as biofertilization mechanisms. Siderophores, which are released by PGPRs, bind Fe³⁺ from the soil and carry it back to the PGPRs cell, where it will be changed into a form that is useful for plant and microbial growth (80,81). Siderophores are produced in substantial amounts in soil, which enables PGPRs to take all the iron required for their growth and deny phytopathogen bacteria access to it. Numerous authors have demonstrated the generation of siderophores by Bacillus and Pseudomonas species (82,83,84,85,60). Numerous studies have demonstrated that the sidorophores produced by the PGPR have a substantial impact on how plants absorb different metals, such as Fe, Zn, and Cu (86,87,88). The bacterial siderophores are able to sequester iron from the rhizosphere, which prevents pathogenic fungi from growing and has a negative impact on plant nutrition. Pseudomonas fluorescens' pseudobactin and pyoverdin siderophores have a definite function in the management of Fusarium species (89,90). Pseudomonas species produced sidorophores that were used in the biocontrol of plant infections like Aspergillus niger (91). These substances are crucial in promoting plant growth, and some plants can directly absorb iron from Pseudomonas siderophore (92). Seed inoculation with PGPRs that produce siderophores enhances plant development and boosts chlorophyll content (93). There is potential for using PGPRs that produce siderophores in agriculture as biocontrol agents and microorganisms that stimulate plant growth.

2.2.2 Chitinase Production

Insoluble linear polymers of ß (1,4) N-acetylglucosamine serve as the main structural constituents of the cell walls of many fungi, insect exoskeletons, and crustacean shells. Chitinase hydrolyzes these polymers to form cell walls (94). Bacillus cereus, which inhibits *Botrytis elliptica* growth, and *Bacillus* sp. S7LiBe, which inhibits B. cinerea growth, are two microbes that generate this enzyme and act as biological control agents (94,95). Multiple investigations have shown that chitinases contribute to antifungal effect and can increase the activity of Bacillus spinsecticidal (96). The majority of Bacillus spp. have significant chitinase activity (97). Some investigations have found a connection between the capacity of Bacillus species and Pseudomonas species to prevent the mycelial growth of Fusarium oxysporum and Fusarium solani. The ability of the chitinase-producing actinomycete (Streptomyces vinaceusdrappus S5MW2) to promote tomato development and act as a biocontrol agent against Rhizoctonia solani. Chitinases are created as a biocontrol method to stop phytopathogenic fungi's spores from germinating (98). In a greenhouse experiment, chitin addition and usage of the bacterial strain S5MW2 significantly increased tomato plant growth and reduced Rhizoctonia solani disease symptoms. Bacteria that produce chitinase have been identified as potential biocontrol agents (99,100).

2.3 Competition for Space and Nutrients

Many and various microorganisms interact in soil, or more specifically in the rhizosphere, and plant roots, which are frequently the only sources of nutrients, are frequently insufficient to support the whole microbial flora. These bacteria compete with one another for all nutrient elements in order to maintain their levels of growth, development, and activity. Competition includes the use of or control over access to resources, such as food, space, or any other finite resource (101). Hattori describes how PGPRs can outcompete harmful organisms by receiving the bulk of nutrients and occupying favorable niches, which causes them to make up a significant fraction of the rhizosphere-rhizoplane population. By competing with them for resources like nitrogen, carbon, or macro or micronutrients, antagonist PGPRs can stop the growth of a variety of phytopathogenic diseases (102). The battle for iron is one particular example of nutritional rivalry. In an effort to live, bacteria release siderophores, as previously mentioned, depriving phytopathogenic agents of one of their growth factors (71). Pseudomonas bacteria may effectively chelate substances

by utilizing siderophores made by other bacteria (103). Plant illness can be lessened by enabling helpful bacteria to colonize a substantial portion of a plant's roots. Because of this colonization, there are fewer places where harmful germs can exist and grow. Two ways for which biocontrol agents can prevent plant growth have been proposed: competition for the substrate (104) and siderophores' competition for iron (105,106).

2.3.1 Antibiosis

Antibiosis, a process where chemicals with antifungal and/or antibiotic properties are produced, inhibits pathogens (103). For instance, among the metabolic products that PGPR produced that possessed bioactive qualities were lytic enzymes (chitinase, protease, glucanase, etc.), antimicrobial proteins or peptides, polyketides, phenolic compounds, bio-surfactants, etc (22). Bacillus and Pseudomonas produce the antibiotics fengycin A and B, iturin A, mycosubtilin, bacillomycin D and pyochelin to attack aflatoxigenic fungus (107). Several publications have discussed PGPR's capacity to produce volatile substances such as ammonia, hydrogen cyanide, acetoin, and 2,3-butanediol. An further class of antibiotic compounds is volatile compounds (108,109,110). Fusarium oxysporum's mycelial development and spore germination were greatly prevented by the volatile chemical produced by bacteria. *Gaeumannomyces graminis* var. tritici "take-all" of wheat was once more suppressed by a cyanide-negative mutant of CHA0 that was complemented by the hcn+ genes (111). Pseudomonas is able to produce significant amounts of the enzymes chitinase and 1,3-glucanase, which degrade the chitin and glucan present in the cell walls of phytopathogenic fungus. Moreover, Pseudomonas can make metabolites that are antifungal. A cyanogenic Pseudomonas bacterium called P. fluorescens CHA0 was engaged in the Competition for nutrients, particularly for carbon and suggested as one of the mechanisms creating the fungistatic effect, which is characterised by the inhibition of spore germination in soil (112). bacteria can parasitize and degrade the spores or hyphae of fungal infections by the creation of a number of enzymes (113). The infection process depends on the generation of hydrolytic enzymes during the early hostpathogen interaction (114). Control of several plant diseases, particularly fungi (111,115). Rhizosphere bacteria produce this chemical, which gives plants a generalized resistance to a variety of diseases (116). One such strain that creates antimicrobial metabolites with a wide range of antifungal activity is P. protegens CHAO, a strain of Pseudomonas fluorescens that produces the antibiotic metabolites 2,4diacetylphloroglucinol (DAPG), pyoluteorin (PLT), and pyrrolnitrin (PRN) (117). One example of how Weller classified several strains of P. fluorescens in which the production of chemicals like phenazines and DAPG is directly related to an antagonistic activity against various pathogens is the antagonistic activity of P. fluorescens 2-79 and CHAO on Gaeumannomyces graminis. Pseudomonas PsJN has also been shown by Siddiqui to reduce the *Botrytis cinerea* caused tomato disease(118,119). The strains of *Pseudomonas fluorescens* PF1, FP7, and PB2 have been shown to inhibit the germination of Rhizoctonia solani sclerotia.

2.3.2 Induced Systemic Resistance

The stimulation of plant defence mechanisms is a component of the PGPRs-plant interaction involved in the control of infections. The term "induced systemic resistance" (ISR) refers to this occurrence, which increases the host's resistance to pathogen aggressiveness in the future. ISR consequently has phenotypic characteristics with RSA, which is brought on by phytopathogenic agents. ISR is as efficient against several pathogens as SAR, however it differs from SAR in that the bacterium that causes PGPR colonises roots and does not manifest any symptoms on the host plant (120,121). When PGPRs interact with plants, they cause structural and physiological changes that lead to the production of molecules involved in plant defense mechanisms. Bacterial lipopolysaccharides, siderophores, and salicylic acid (SA) are found to be the major determinants of ISR. Systemic resistance may be induced by various microorganisms, Gram-positive bacteria such as Bacillus pumilus, or Gramnegative bacteria belonging to the genus Pseudomonas (fluorescens, putida, aeruginosa) (122). Arabidopsis seedlings exposed in divided Petri dishes to PGPR B. subtilis GB03 and B. amyloliquefaciens IN937a for 10 days developed significantly less symptomatic leaves 24 h after inoculation with the soft rot-causing pathogen Erwinia carotovora ssp. carotovora, this suggests that the volatile compounds play an important role in the induction of plant resistance (120,108). In an experiment carried out in the field, Jetiyanon reported that a combination of B. amyloliquefaciens strain IN937a and B. pumilus strain IN937b induced systemic resistance against southern blight of tomato, caused by Sclerotium rolfsii, anthracnose of long cayenne pepper, caused by *Colletotrichum gloeosporioides* (123) and mosaic disease of cucumber (124), P. fluorescens 63-28inoculated pea roots produced more chitinase at the Fusarium oxysporum penetration site. The suppression of Rhizoctonia solani by Pseudomonas fluorescens through the induction of plant resistance system has also been documented. P. *fluorescens* spp. release an insecticidal toxin and make plants more resistant to aphid assault. It can be said that PGPR will be of great interest, especially to protect plants and avoid issues encountered when pesticides fail to control pathogen populations that have developed resistance (125). PGPR belonging to Pseudomonas spp. are commercially exploited to protect plants by inducing their systemic resistance against various pests and diseases (126).

2.3.3 Agricultural Application

The use of PGPB as agricultural inoculants is a possible substitute based on these bacteria's capacity for biofertilization and biocontrol. The application and fate of inoculants on fieldgrown crops must be thoroughly checked in order to ensure that inoculants can boost yields in a way that can be demonstrably shown (127). Understanding the factors that control and direct the generation of bioactive compounds is vital for increasing the quantity and consistency of PGPB activity. It may be required to first look at how these parameters affect PGPB activities in vitro in order to optimize these activities in vivo or to develop bioactive metabolites. Environmental factors such as soil type, pH, plant surface, and climatic conditions are known to have an effect on how well PGPR operates. Abiotic factors are also the main determinants of PGPR effectiveness (86). The pH and incubation temperature have an effect on antagonist activity, Several authors have discussed how different abiotic factors, like temperature and nutritional components, affect the generation of antifungal chemicals by biocontrol agents (128). According to Naik and Sakthivel, bacteria that are related to pseudomonas may use a range of carbon sources as a substrate and can adapt to different conditions (129). A wide range of

abiotic factors, including glucose, Fe+3, Zn+2, Cu+2, and Mo+2, influence the production of antifungal compounds. Due to the presence of Fe+ 3 and sucrose, diacetyl-phloroglucinol (DAPG) was formed in P at a greater rate. In contrast to P. fluorescens Pf-5 and CHA0, glucose increases the development of P. fluorescens strain F113 (130). One illustration of how nutrients and environmental factors might impact the antifungal activity of P. *cepacia* is the carbon supply, which promotes antagonist action and inhibits the formation of the spores (128). Temperature has a significant impact on bacteria's capacity to produce antifungal compounds. Another element that may affect the bioactive potential of bacterial inoculants is their ability to adapt to the soil's conditions and compete with native species (127). By using mathematical modelling and computer-based simulation, Strigul and Kravchenko aimed to evaluate microbial inoculants in the rhizosphere and the impact of different abiotic and biotic stress on PGPBs survival and activity (127). Strigul and Kravchenko have shown that the most important factor affecting the survival of inoculants was the struggle for few resources between the introduced population and the native microorganisms (131). The survival of the inoculants was also influenced by the inoculated bacteria's capacity to utilise specific elements found in the exudates from the roots of the host plant. Understanding the many pathways involved in promoting plant development and squelching disease is essential for the selection and application of suitable biocontrol strains for sustainable agriculture (132). As was previously said, in order to ensure an improvement in field-grown crop yields, due consideration must be given to the choice of PGPBs inoculants as well as the impact of the various environmental conditions. The application method (seed treatment, postharvest treatment, foliar spray, etc.) for the bacterial inoculant may also affect how active it is.

3. Conclusion and Future Prospects

Use of PGPBs in agriculture is a workable solution that can enhance plant health and performance. One can boost food production and meet increased demand by knowing how to protect plants and encourage their growth. The PGPB findings indicate a cultural change towards "bio-agriculture," which is beneficial to both the national economy and human health. The usage of PGPBs in agriculture can serve as a feasible alternative to chemical products (such fertilizers and pesticides) that are harmful to the environment and the health of the general people. PGPBS have all been previously reported as having the ability to biofertilize soil, biocontrol phytopathogenic agents and promote plant growth, which encourages their commercial exploitation and usage as inoculants.

References

- Kouassi, M. 2001. La lutte biologique: une alternative viable à l'utilisation des pesticides. Vertigo La revue électronique en sciences de l'environnement, volume 2, Octobre 2001 Issue. http://vertigorevues.org/4101; https://doi.org/10.4000/vertigo4101.
- 2. Thakore, Y. 2006. The biopesticide market for global agricultural use. *Ind Biotechnol* 2:294–208.
- Nyembo, K. L., Useni, S. Y., Mpundu, M. M., Bugeme, M. D., Kasongo, L. E., Baboy, L. L. 2012. Effets des apports des doses variées de fertilisants inorganiques (NPKS et Urée) sur le rendement et la rentabilité économique de nouvelles

variétés de Zeamays L à Lubumbashi, Sud-Est de la RD Congo. *J Appl Biosc* 59:4286–4296.

- Faessel, L., Gomy, C., Nassr, N., Tostivint, C., Hipper, C. and Dechanteloup, A. 2014. Produits de stimulation en agriculture visantà améliorer les fonctionnalités biologiques des sols et des plantesÉtude des connaissances disponibles et recommandations stratégiques. BIO&RITTMO, 156 pp.
- De Salamon, D. I. G., Hynes, R. K., Nelson, L. M. 2006. Role of cytokinins in plant growth promotion by rhizosphère bacteria. In: Siddiqui AA (ed) PGPR: biocontrol and biofertilization. *Springer*, Dordrecht, pp 173–195.
- 6. Bensidhoum, L., Nabti, E., Tabli, N., Kupferschmied, P., Weiss, A., Rothballer, M., Schmid, M., Keel, C., Hartmann, A. 2016. Heavy metal tolerant Pseudomonas protegens isolates from agricultural well water in northeastern Algeria with plant growth promoting, insecticidal and antifungal activities. *Eur J Soil Biol* 75:38–46.
- 7. Gray, E. J., Smith, D. L. 2005. Intracellular and extracellular PGPR: commonalities and distinctions in the plant-bacterium signalling processes. *Soil Biol Biochem* 37:395-412.
- Tokala, R. K., Strap, J. L., Jung, C. M., Crawford, D. L., Salove, H., Deobald, L. A., Bailey, F. J., Morra, M. J. 2002. Novel plant microbe rhizosphere interaction involving S lydicus WYEC108 and the pea plant (Pisum sativum). *Appl Environ Microbiol* 68:2161–2171.
- 9. Yadav, A. N., Verma, P., Kour, D., Rana, K. L., Kumar, V., Singh, B. 2017b. Plant microbiomes and its beneficial multifunctional plant growth promoting attributes. Int J E n v i r o n S c i N a t R e s o u r 3 : 1 - 8. https://doi.org/10.19080/IJESNR.2017.03.555601
- Yadav, A. N., Kumar, V., Prasad, R., Saxena, A. K., Dhaliwal, H. S. 2018a. Microbiome in crops: diversity, distribution and potential role in crops improvements. In: Prasad R, Gill SS, Tuteja N (eds) Crop improvement through microbial biotechnology. *Elsevier, Amsterdam*, pp 305–332.
- 11. Rifat, H., Safdar, A., Ummay, A., Rabia, K., Iftikhar, A. 2010. Soil beneficial bacteria and their role in plant growth promotion. *Ann Microbiol* 60:579–598.
- 12. Yadav, A. N., Verma, P., Singh, B., Chauhan, V. S., Suman, A., Saxena, A. K. 2017c. Plant growth promoting bacteria: biodiversity and multifunctional attributes for sustainable agriculture. *Adv Biotechnol Microbiol* 5:1–16.
- 13. Compant, S., Duffy, D., Nowak, J., Clément, C., Ait Barka, E. 2005. Use of plant growth promoting Bacteria for biocontrol of plant diseases: principles, mechanisms of action, and future prospects. *Appl Environ Microb* 71:4951–4959.
- Vacheron, J., Desbrosses, G., Bouffaud, M. L., Touraine, B., Moenne-Loccoz, Y., Muller, D., Legendre, L., Wisniewski-Dye, F., Prigent-Combaret, C. 2013. Plant growth promoting rhizobacteria and root system functioning. *Front Plant Sci* 4:1–19.

- 15. Kour, D., Rana, K. L., Yadav, N., Yadav, A. N., Kumar, A., Meena, V. S. 2019c. Rhizospheric microbiomes: biodiversity, mechanisms of plant growth promotion, and biotechnological applications for sustainable agriculture. In: Kumar A, Meena VS (eds) Plant growth promoting Rhizobacteria for agricultural sustainability: from theory to practices. Springer Singapore, Singapore, pp 19–65. https://doi.org/10.1007/978-981-13-7553-8_2
- 16. Verma, P., Yadav, A. N., Khannam, K. S., Panjiar, N., Kumar, S., Saxena, A. K. 2015. Assessment of genetic diversity and plant growth promoting attributes of psychrotolerant bacteria allied with wheat (*Triticum aestivum*) from the northern hills zone of India. *Ann Microbiol* 65:1885–1899.
- 17. O'sullivan, D. J., O'gara, F. 1992. Traits of fluorescent Pseudomonas spp. involved in suppression of plant root pathogens. *Microbiol Rev* 56:662–676.
- 18. Patten, C. L., Glick, B. R. 2002. Regulation of indole acetic acid production in Pseudomonas putida GR12-2 by tryptophan and the stationary-phase sigma factor. *RpoS CanJ Microbiol* 48:635–642.
- 19. Glick, B. R. 1995. The enhancement of plant growth by freeliving bacteria. *Can J Microbial* 41:109–117.
- Anton, H., Prevost, D. 2006. Ecology of plant growth promoting rhizobacteria. In: Siddiqui ZA (ed) PGPR: biocontrol and biofertilization. *Springer Netherlands*, Dordrecht, pp 1–38. https:// doi.org/10.1007/1-4020-4152-7_1.
- 21. Kaymak, H. 2010. Potential of PGPR in agricultural innovations. In: Maheshwari DK (ed) Plant growth and health promoting bacteria. *Springer*, Berlin/Heidelberg, pp 45–79.
- 22. Fernando, W. G. D., Nakkeeran, S., Zhang, Y. 2005. Biosynthesis of antibiotics by PGPR and its relation in biocontrol of plant diseases. In: Siddiqui ZA (ed) PGPR: biocontrol and biofertilization. *Springer*, Dordrecht, pp 67–109.
- 23. Fuentes-Ramirez, L. E., Caballero-Mellado, J. 2005. Bacterial biofertilizers. In: Siddiqui ZA (ed) PGPR: biocontrol and biofertilization. *Springer*, Dordrecht, pp 143–172.
- 24. Verma, P., Yadav, A. N., Khannam, K. S., Mishra, S., Kumar, S., Saxena, A. K. 2019. Appraisal of diversity and functional attributes of thermotolerant wheat associated bacteria from the peninsular zone of India. *Saudi J Biol Sci* 26:1882–1895. https://doi.org/10.1016/j.sjbs. 2016.01.042
- 25. Yadav, A. N., Kumar, R., Kumar, S., Kumar, V., Sugitha, T., Singh, B. 2017a. Beneficial microbiomes: biodiversity and potential biotechnological applications for sustainable agriculture and human health. *J Appl Biol Biotechnol* 5:45–57.

- Verma, .P, Yadav, A. N., Khannam, K. S., Kumar, S., Saxena, A. K., Suman, A. 2016. Molecular diversity and multifarious plant growth promoting attributes of Bacilli associated with wheat (Triticum aestivum L.) rhizosphere from six diverse agro-ecological zones of India. J Basic Microbiol 56:44–58
- 27. Yadav, A. N., Gulati, S, Sharma, D., Singh, R. N., Rajawat, M. V. S., Kumar, R. 2019a. Seasonal variations in culturable archaea and their plant growth promoting attributes to predict their role in establishment of vegetation in Rann of Kutch. *Biologia* 74:1031–1043. https://doi.org/10.2478/s11756-019-00259-2.
- 28. Tripathi, M., Munot, H. P., Van Loon, L. C. 2007. Plant responses to plant-growth promoting rhizobacteria. *Eur J Plant Pathol* 119:243–254.
- 29. Somers, E., Vanderleyden, J., Srinivasan, M. 2004. Rhizosphere bacterial signalling: a love parade beneath our feet. *Crit Rev Microbiol* 304:205–240.
- Kumar, Sethi, S., Kumari Sahu, J, Prasad Adhikary, S. 2015. Microbial biofertilizers and their pilotscale production. In: Harzevili FD, Chen H (eds) Microbial biotechnology. Progress and trends. Taylor & Francis Group, Boca Raton, pp 297–315.
- 31. Kour, D., Rana, K. L., Yadav, N., Yadav, A. N., Singh, J., Rastegari, A. A. 2019d. Agriculturally and industrially important fungi: current developments and potential biotechnological applications. In: Yadav AN, Singh S, Mishra S, Gupta A (eds) Recent advancement in white biotechnology through fungi. Volume 2: Perspective for value-added products and environments. *Springer* International Publishing, Cham, pp 1–64. https://doi.org/10.1007/978-3-030-14846-1_1
- Rana, K. L., Kour, D., Sheikh, I., Yadav, N., Yadav, A. N., Kumar, V. 2019. Biodiversity of endophytic fungi from diverse niches and their biotechnological applications. In: Singh BP (ed) Advances in endophytic fungal research: present status and future challenges. *Springer* International P u b l i s h i n g, C h a m, p p 1 0 5 1 4 4. https://doi.org/10.1007/978-3-030-03589-1_6.
- Yadav, A. N., Verma, P., Kumar, S., Kumar, V., Kumar, M., Singh, B. P. 2018b. Actinobacteria from rhizosphere: molecular diversity, distributions and potential biotechnological applications. In: Singh B, Gupta V, Passari A (eds) New and future developments in microbial biotechnology and bioengineering. *Elsevier, Amsterdam*, pp 13–41. https://doi.org/10.1016/B978-0-444-63994-3.00002-3.
- 34. Bashan, Y., Holguin, G., de-Bashan, L. E. 2004. Azospirillumplant relationships: physiological, molecular, agricultural, and environmental advances (1997-2003). *Can J Microbiol* 50:521–577.
- 35. Stamford, N. P., Santos, C. E. R. S., Junior, S. S., Lira Junior, M. A., Figueiredo, M. V. B. 2008. Effect of rhizobia and rock biofertilizers with Acidithiobacillus on cowpea nodulation and nutrients uptake in a tableland soil. *World J Microbiol Biotechnol* 24:1857–1865.

- 36. Chabot, R., Antoun, H., Cescas, M. P. 1996. Growth promotion of maize and lettuce by phosphatesolubilizing Rhizobium leguminosarum by phaseoli. *Plant Soil* 184:311–321.
- 37. Raja, P., Una, S., Gopal, H., Govindarajan, K. 2006. Impact of bio-inoculants consortium on rice root exudates, biological nitrogen fixation and plant growth. *J Biol Sci* 6:815–823.
- 38. Kour, D., Rana, K. L., Kumar, A., Rastegari, A. A., Yadav, N., Yadav, A. N. 2019a. Extremophiles for hydrolytic enzymes productions: biodiversity and potential biotechnological applications. In: Molina G, Gupta VK, Singh BN, Gathergood N (eds) Bioprocessing for biomolecules production. Wiley, Newark, pp 321–372.
- 39. Chet, I., Ordentlich, A., Shapira, R., Oppenheim, A. 1990. Mechanisms of biocontrol of soil-borne plant pathogens by rhizobacteria. *Plant Soil* 129:85–92.
- 40. Kobayashi, D. Y., Reedy, R. M., Bick, J. A., Oudemans, P. V. 2002. Characterization of a chitinase gene from *Stenotrophomonas maltophilia* strain 34S1 and its involvement in biological control. Appl Environ Microbiol 68:1047–1054.
- 41. Ljungdahl, L. G., Eriksson, K. E. 1985. Ecology of microbial cellulose degradation. *Adv Microb Ecol* 8:237–299.
- 42. Sindhu, S. S., Dadarwal, K. R. 2001. Chitinolytic and cellulolytic Pseudomonas sp antagonistic to fungal pathogens enhances nodulation by Mesorhizobium sp Cicer in chickpea. *Microbiol Res* 156:353–358.
- Petit, J. and Jobin, P. 2005. La fertilisation organique des cultures. Les bases Fédération d'agriculture biologique du Québec Bibliothèque national de Canada, 48p Polacco JC (1977) Is nickel a universal component of plant ureases? *Plant Sci Lett* 10:249–255.
- 44. Twisha, P., Desai, P. B. 2014. Study on Rhizospheric microflora of wild and transgenic varieties of Gossypium species in monsoon. *Res J Recent Sci* 3:42–51.
- 45. Lian, L. H., Tian, B. Y., Xiong, R., Zhu, M. Z., Xu, J., Zhang, D. Q. 2007. Proteases from Bacillus: a new insight into the mechanism of action for rhizobacterial suppression of nematode populations. *Lett Appl Microbiol* 45:262–269.
- 46. Dunne, C., Crowley, J. J., Moenne-Loccoz, Y., Dowling, D. N., De Bruijin, F. J., O'Gara, F. 1997. Biological control of *Pythium ultimum* by *Stenotrophomonas maltophilia* W81 is mediated by an extracellular proteolytic activity. *Microbiology* 143:3921–3931.
- 47. Mansour, F. A., Aldesuquy, H. S., Hamedo, H. A. 1994. Studies on plant growth regulators and enzymes production by some Bacteria. *Qatar Univ Sci J* 14:281–288.
- Martinez-Salgado, M. M., Gutiérrez-Romero, V., Jannsens, M., Ortega-Blu, R. 2010. Biological soil quality indicators: a review. In: Mendez-Vilas A (ed) Current research, technology and education topics in applied microbiology and microbial biotechnology. *Formatex, Badajoz*, pp 319–328.

- 49. Kumar, D. S., Varma, A. 2011. Role of enzymes in maintaining soil health. In: Shukla G, Varma A (eds) Soil enzymology. *Springer*, Berlin/Heidelberg, pp 25–42.
- 50. Mobley, H. L. T., Hausinger, R. P. 1989. Microbial urease: significance, regulation and molecular characterization. *Microbiol Rev* 53:85–108.
- 51. Pujic, P., Normand, P. 2009. La symbiose racinaire entre la bactérie Frankia et les plantes actinorhiziennes. *Biofutur* 298:26–29.
- 52. Doty, S. L. 2011. Nitrogen-fixing endophytic bacteria for improved plant growth. In: Maheshwari DK (ed) Bacteria in agrobiology: plant growth responses. *Springer*, Berlin/Heidelberg, pp 183–199.
- 53. Townsend, A. R., Howarth, R. W. 2010. Fixing the global nitrogen problem. *SciAm* 302(2):64–71.
- 54. Okon, Y., Kapulnik, Y. 1986. Development and function of Azospirillum-inoculated roots. *Plant Soil* 90:3–16.
- 55. Suman, A., Yadav, A. N., Verma, P. 2016. Endophytic microbes in crops: diversity and beneficial impact for sustainable agriculture. In: Singh D, Abhilash P, Prabha R (eds) Microbial inoculants in sustainable agricultural productivity. Research perspectives. *Springer*, New Delhi, pp 117–143. https://doi.org/10.1007/978-81-322-2647-5_7.
- 56. Bensidhoum, L., Nabti, E. 2019. Plant growth-promoting Bacteria for improving crops under saline conditions. In: Giri B, Varma A (eds) Microorganisms in saline environments: strategies and functions. Soil biology 56. Springer Nature, Cham. https://doi.org/10.1007/978-3-030-18975-4_14.
- 57. Vessey, J. K. 2003. Plant growth promoting rhizobacteria as biofertilizers. *Plant Soil* 255:571–586.
- 58. Dobereiner, J. 1997. Biolgical nitrogen fixation in the tropics: social and economic contributions. *Soil Biol Biochem* 29:771–774.
- 59. Naqqash, T., Hameed, S., Imran, A., Hanif, M. K., Majeed, A., van Elsas, J. D. 2016. Differential response of potato toward inoculation with taxonomically diverse plant growth promoting rhizobacteria. *Front Plant Sci* 7:144.
- 60. Przemieniecki, S. W., Kurowski, T. P., Karwowska, A. 2015. Plant growth promoting potential of Pseudomonas sp. Sp0113 isolated from potable water from a closed water well. *Arch Biol Sci Belgrade* 67:663–673.
- 61. Nabti, E. 2007. Restauration de la croissance de Azospirillum brasilense et de Blé dur et leur osmoprotection par Ulva lactuca enMilieux Salés. Thèse de Doctorat en Science Biologique Université Abderrahmane Mira, Faculté des sciences de la nature et de la vie, Bejaia, 147p.
- 62. Richardson, A. E. 2001. Prospects for using soil microorganisms to improve the acquisition of phosphorus by plants. *Aust J Plant Physiol* 28:897–906.

- 63. Qureshi, M. A., Ahmad, Z. A., Akhtar, N., Iqbal, A., Mujeeb, F., Shakir, M. A. 2012. Role of phosphate solubilizing bacteria (psb) in enhancing P availability and promoting cotton growth. *J Anim Plant Sci* 22:204–210.
- 64. Kour, D., Rana, K. L., Yadav, A. N., Yadav, N., Kumar, V., Kumar, A. 2019b. Drought-tolerant phosphorus-solubilizing microbes: biodiversity and biotechnological applications for alleviation of drought stress in plants. In: Sayyed RZ, Arora NK, Reddy MS (eds) Plant growth promoting rhizobacteria for sustainable stress management: Volume 1: Rhizobacteria in abiotic stress management. *Springer* Singapore, Singapore, pp 255–308. https://doi.org/10.1007/978-981-13-6536-2_13.
- 65. Yadav, A. N., Verma, P., Kumar, V., Sangwan, P., Mishra, S., Panjiar, N. 2018c. Biodiversity of the genus Penicillium in different habitats. In: Gupta VK, Rodriguez-Couto S (eds) New and future developments in microbial biotechnology and bioengineering. Penicillium system properties and applications. *Elsevier, Amsterdam,* pp 3–18. https://doi.org/10.1016/B978-0-444-63501-3.00001-6.
- 66. Glick, B. 2012. Plant growth-promoting bacteria: mechanisms and applications. *Scientifica* 2012:1–15.
- 67. Oteino, N., Lally, R. D., Kiwanuka, S., Andrew, L., Ryan, D., Germaine, K. J., Dowling, D. N. 2015. Plant growth promotion induced by phosphate solubilizing endophytic Pseudomonas isolates. *Front Microbiol* 6:1–9.
- 68. Trivedi, P., Sa. T. 2008. *Pseudomonas corrugata* (NRRL B-30409) mutants increased phosphate solubilization, organic acid production, and plant growth at lower temperatures. *Curr Microbiol* 56:140–144
- 69. Rodríguez, H., Fraga, R. 1999. Phosphate solubilizing bacteria and their role in plantgrowth promotion. *Biotechnol Adv* 17:319–339.
- 70. Elkoca, E., Kantar ,F., Sahin, F. 2008. Influence of nitrogen fixing and phosphorus solubilizing bacteria on the nodulation, plant growth, and yield of chickpea. *J Plant Nutr* 31:157–171.
- 71. Pal, K. K., Gardener, B. M. S. 2006. Biological control of plant pathogens. *Plant Health Instr* 2:1–25.
- 72. Niranjan, R. S., Shetty, H. S., Reddy, M. S. 2005. Plant Growth Promoting Rhizobacteria: potential green alternative for plant productivity. In: Siddiqui ZA (ed) PGPR: biocontrol and biofertilization. Springer, Dordrecht, pp 197–216.
- 73. Lee, M., Qi, M., Yang, Y. 2001. A novel jasmonic acidinducible rice myb gene associates with fungal infection and host cell death. *Mol Plant-Microbe Interact* 14:527-535.
- 74. Marschner, P., Timonen, S. 2006. Bacterial community composition and activity in rhizosphere of roots colonized by arbuscular mycorrhizal fungi. In: Mukerji KG, Manoharachary C, Singh J (eds) Microbial activity in the rhizosphere. Soil Biology 7. *Springer*, New York, pp 139–154.

- 75. Jamali, F., Shi-Tehrani, A., Lutz, M. P., Maurhofer, M. 2009. Influence of host plant génotype, presence of a pathogen, and coinoculation with *Pseudomonas fluorescens* strains on the rhizosphere expression of hydrogen cyanide- and 2,4-Diacetylphloroglucinol biosynthesis genes in *P. fluorescens* biocontrol strain CHAO. *Microb Ecol* 57:267–275.
- 76. Calvo, P., Ormeño-Orrillo, E., Martínez-Romero, E., Zúñiga, D. 2010. Characterization of Bacillus isolates of potato rhizosphere from andean soils of Peru and their potential PGPR characteristics. *Braz J Microbiol* 41:899–906.
- 77. Bensidhoum, L., Rai, A., Tabli, N., Kahouadji, N., Khaber, M., Nabti, E. 2015. Biological control of *Botrytis cinerea* by Bacillus sp. strain S7LiBe under abiotic stress. *Int J Sci Res Sci Technol* 6:07–14.
- 78. Tabli, N., Rai, A., Bensidhoum, L., Palmieri, G., Gogliettino, M., Cocca, E., Consiglio, C., Cillo, F., Bubici, G., Nabti, E. 2018. Plant growth promoting and inducible antifungal activities of irrigation well water-bacteria. *Biol Control* 117:78–86.
- 79. Manteau, S., Abouna, S., Lambert, B., Legendre, L. 2003. Differential regulation by ambient pH of putative virulence factor secretion by the phytopathogenic fungus Botrytis cinerea. *FEMS Microbiol Ecol* 43:359–366.
- Beneduzi, A., Passaglia, L. M. P. 2011. Genetic and phenotypic diversity of plant growth promoting bacilli. In: Maheshwari DK (ed) Bacteria in agrobiology: plant growth responses. *Springer*, Berlin/Heidelberg, pp 1–20.
- 81. Leong, J. 1986. Siderophores: their biochemistry and possible role in the biocontrol of plant pathogens. *Annu Rev Phytopathol* 24:187–209.
- 82. Meyer, J. M., Abdallah, M. A. 1978. The fluorescent pigment of *Pseudomonas fluorescens:* biosynthesis, purification and physicochemical properties. *J Gen Microbiol* 107:319–328.
- 83. Zahir, Z. A., Arshad, M., Frankenberger, W. T. J. 2004. Plant growth promoting rhizobacteria: application and perspectives in agriculture. *Adv Agro* 81:97–198.
- 84. Mezaache, S. 2012. Localisation des déterminants de la suppression de quelques souches de pseudomonas isolées de la rhizosphère de la pomme de terre. Thèse de Doctorat en Sciences, Université de Ferhat Abbas de Sétif, Faculté des Sciences de la Nature et de la Vie, Sétif, 221p.
- 85. Ji, S. H., Gururani, M. A., Chul, C. S. 2014. Isolation and characterization of plant growth promoting entophytic diazotrophic bacteria from Korean rice cultivars. *Microbiol Res* 169:83–98.
- 86. Egamberdieva, D. 2012. The management of soil quality and plant productivity in stressed environment with rhizobacteria. In: Maheshwari DK (ed) Bacteria in agrobiology: stress management. Springer, Heidelberg/Dordrecht/London/NewYork, pp 27-40.
- 87. Dimkpa, C., Svatos, A., Merten, D., Büchel, G., Kothe, E. 2008. Hydroxamate siderophores produced by *Streptomyces acidiscabies* E13 bind nickel and promote growth in cowpea (*Vigna unguiculata* L.) under nickel stress. *Can J Microbiol* 54:163–172.

- 88. Gururani, M. A., Upadhyaya, C. P., Baskar, V., Venkatesh, J., Nookaraju, A., Park, S. W. 2012. Plant growth promoting rhizobacteria enhance abiotic stress tolerance in Solanum tuberosum through inducing changes in the expression of ROS-scavenging enzymes and improved photosynthetic performance. *J Plant Growth Regul* 32:245–258.
- Alabouvette, C., Schippers, B., Lemanceau, P., Bakker, P. A. H. M. 1998. Biological control of Fusarium wilts toward development of commercial products. In: Boland GC, Kuykendall LD (eds) Plantmicrobe interactions and biological control. Marcel Dekker, New York, pp 15–36.
- 90. Chincholkar, S. B., Chaudhari, B. L., Rane, M. R. 2007. Microbial siderophore: a state of art. *Soil Biol* 12:233–242.
- 91. Sindhu, S. S., Dua, S., Verma, M. K., Khandelwal, A. 2010. Growth promotion of legumes by inoculation of rhizosphere bacteria. In: Khan MS, Musarat J, Zaidi A (eds) Microbes for legume improvement. *Springer*, Vienna, pp 195–235.
- 92. Bar-Ness, E., Chen, Y., Hader, Y., Marschner, H., Romheld, V. 1991. Siderophores of *Pseudomonas putida* as an Iron source for dicot and monocot plants. In: Chen Y, Hader Y (eds) Iron nutrition and interaction in plants. Kluwer Academic Publishers, Dordrecht, *The Netherlands*, pp 271–281.
- 93. Sharma, A., Johri, B. N. 2003. Combat of iron-deprivation through a plant growth promoting fluorescent Pseudomonas strain GRP3A in mung bean (*Vigna radiate* L. Wilzeck). *Microbiol Res* 158:77–81.
- 94. Bhushan, B., Hoondal, G. S. 1998. Isolation, purification and properties of a thermostable chitinase from an alkalophilic Bacillus sp. BG-11. *Biotechnol Lett* 20:157–159.
- Huang, X. D., El-Alawai, Y., Gurska, J., Glick, B. R., Greenberg, B. M. 2005. A multi-process phytoremediation system for decontamination of persistent total petroleumhydrocarbons (TPHs) from soils. *Microchemistry* J81:139–147.
- 96. Quan, C. S., Wang, X., Fan, S. D. 2010. Antifungal compounds of plant growth promoting rhizobacteria and its action mode. In: Maheshwari DK (ed) Plant growth and health promoting bacteria. *Springer*, Berlin/Heidelberg, pp 117–156.
- 97. Quecine, M. C., Araujo, W. L., Marcon, J., Gail, C. S., Azevedo, J. L., Pizzirani-Kleiner, A. A. 2008. Chitinolytic activity of endophytic Streptomyces and potential for biocontrol. *Lett Appl Microbiol* 47:486–491.
- 98. Nandakumar, R., Babu, S., Radjacommare, R., Raguchander, T., Samiyappan, R. 2002. *Pseudomonas fluorescens* mediated antifungal activity against Rhizoctonia solani causing sheath blight in rice. *Phytopathol Mediterr* 41:109–119.
- 99. Ordentlich, A., Elad, Y., Cher, I. 1988. The role of chitinase of *Serrattia marcescens* in biocontrol of *Sclerotium rolfsii*. *Phytopathology* 78:84–88.
- 100. Inbar, J., Chet, I. 1991. Evidence that chitinase produced by *Aeromonas caviae* is involved in the biological control of soilborne plant pathogens by this bacteria. *Soil Biol Biochem* 23:973–978.

- 101. Widen, B., Cronberg, N., Widen, M. 1994. Genotypic diversity, molecular markers and spatial distribution of genets in clonal plants, a literature survey. *Folia Geobot* 29:245–263.
- 102. Elad, Y., Stewart, A. 2007. Microbial control of Botrytis spp. In: Elad Y, Willizmson B, Tudzynski P, Delen N (eds) Botrytis: biology, pathology and control. *Springer*, Dordrecht, pp 223–241.
- 103. Johansson, P. M. 2003. Biocontrol of Fusarium in wheat -introducing bacteria to a system of complex interactions. Doctoral thesis, Swedish University of Agricultural Sciences Suède, p 74.
- 104. Couteaudier, Y., Alabouvette, C. 1990. Quantitative comparison of *Fusarium oxysporum* competitiveness in relation to carbon utilization. *FEMS Microbiol Lett* 74(4):261–267.
- 105. Raaijmakers, J. M., Leeman, M., Van Oorschot, M. M. P., van der Sluis, I., Schippers, B., Bakker, P. A. H. M. 1995. Doseresponse relationships in biological control of Fusarium wilt of radish by Pseudomonas spp. *Phytopathology* 85(10):1075–1081.
- 106. Pandey, S., Ghosh, P. K., Ghosh, S., De, T. K., Maiti, T. K. 2013. Role of heavy metal resistant Ochrobactrum sp. and Bacillus spp. strains in bioremediation of a rice cultivar and their PGPR like activities. *J Microbiol* 51(1):11–17.
- 107. Ghahfarokhi, M. S., Kalantari, S., Razzaghi-Abyaneh, M. 2013. Terrestrial bacteria from agricultural soils: versatile weapons against aflatoxigenic fungi0. In: Razzaghi-Abyaneh M (Ed) Aflatoxins. Recent advances and future prospects. *InTech*, pp 23–39.
- Ryu, C. M., Farag, M. A., Hu, C. H., Reddy, M. S., Wei, H. X., Paré, P. W., Kloepper, J. W. 2003. Bacterial volatiles promote growth in Arabidopsis. *Proc Natl Acad Sci* 100:4927–4932.
- 109. Charest, M. H., Beauchamp, C. J., Antoun, H. 2005. Effects of the humic substances of de-inking paper sludge on the antagonism between two compost bacteria and *Pythium ultimum. FEMS Microbiol Ecol* 52:219–227.
- 110. Wani, P. A., Khan, M. S., Zaidi, A. 2007. Synergistic effects of the inoculation with nitrogen fixing and phosphate-solubilizing rhizobacteria on the performance of field grown chickpea. *J Plant Nutr Soil Sci* 170:283–287.
- 111. Voisard, C., Keel, C., Haas, D., Défago, G. 1989. Cyanide production by *Pseudomonas fluorescens* helps suppress black root rot of tobacco under gnotobiotic conditions. *EMBOJ* 8:351–358.
- 112. Alabouvette, C., Olivain, C., Steinberg, C. 2006. Biological control of plant diseases: the European situation. *Eur J Plant Pathol* 114:329–341.
- 113. Whipps, J. M. 2001. Microbial interaction and biocontrol in the rhizosphere. *J Exp Bot* 52:487–511.

- 114. Baarlen, V. P., Staats, M., Kan, V. J. 2004. Induction of programmed cell death in lily by the fungual pathogen *Botrytis elliptica. Mol Plant Pathol* 5:559–574.
- 115. Blumer, C., Haas, D. 2000. Mechanism, regulation, and ecological role of bacterial cyanide biosynthesis. *Arch Microbiol* 173:170–177.
- 116. Kumar, N. A., Vanlalzarzova, B., Sridhar, S., Baluswami, M. 2012. Effect of liquid seaweed fertilizer of Sargassum wightii Grev on the growth andbiochemical content of green gram (*Vigna radiata* (L) R Wilczek). *Rec Res Sci Technol* 4:40–45.
- 117. Haas, D., Keel, C. 2003. Regulation of antibiotic production in root-colonizing Pseudomonas spp and relevance for biological control of plant disease. *Annu Rev Phytopathol* 41:117–153.
- 118. Weller, D. M. 2007. Pseudomonas biocontrol agents of soilborne pathogens: looking back over 30 years. *Phytopathology* 97:250–256.
- 119. Siddiqui, Z. A. 2006. PGPR: prospective biocontrol agents of plant pathogens. In: Siddiqui ZA (ed) PGPR: biocontrôle and biofertilization. *Springer*, Pays-Bas, pp 111–142.
- 120. Borriss, R. 2011. Use of plant-associated bacillus strains as biofertilizers and biocontrol agents in agriculture. In: Maheshwari DK (ed) Bacteria in agrobiology: plant growth responses. *Springer*, Berlin/Heidelberg, pp 41–76.
- 121. Van, Loon, L. C., Bakker, P. A. H. M., Pieterse, C. M. J. 1998. Systemic resistance induced by rhizosphere bacteria. *Annu Rev Phytopathol* 36:453–483.
- 122. Jourdan, E., Ongena, M., Thonar, P. 2008. Caractéristiques moléculaires de l'immunité des plantes induite par les rhizobactéries non pathogènes. *Biotechnol Agron Soc Environ* 4:437–449.
- 123. Jetiyanon, K., Fowler, W. D., Kloepper, J. W. 2003. Broadspectrum protection against several pathogens by PGPR mixtures under field conditions in Thailand. *Plant Dis* 87:1390–1394.
- 124. Benhamou, N., Kloepper, J. W., Quadt-Hallmann, A., Tuzun, S. 1996. Induction of defense related ultrastructural modifications in pea root tissues inoculated with endophytic bacteria. *Plant Physiol* 112:919–929.
- 125. Kempster, V. N., Scott, E. S., Davies, K. A. 2002. Evidence for systemic, cross-resistance in white clover (*Trifolium repens*) and annual medic (*Medicago truncatula var truncatula*) induced by biological and chemical agents biocontrol. *Sci Technol* 12:615–623.
- 126. Wang, K. L., Li, H., Ecker, J. R. 2002. Ethylene biosynthesis and signaling networks. *Plant Cell* 14:131–151.
- 127. Cummings, S. P., Orr, C. 2010. The role of plant growth promoting rhizobacteria in sustainable and low-input Graminaceous crop production. In: Maheshwari DK (ed) Plant growth and health promoting bacteria. *Springer*, Berlin/Heidelberg, pp 297–315.

- 128. Upadhyay, R. S., Visintin, L., Jayaswal, R. K. 1991. Environmental factors affecting the antagonism of Pseudomonas cepacia against Trichoderma viride. *Can J Microbiol* 37:880–884.
- 129. Naik, P. R., Sakthivel, N. 2006. Functional characterization of a novel hydrocarbonoclastic Pseudomonas sp strain PUP6 with plant-growth-promoting traits and antifungal potential. *Res Microbiol* 157:538–546.
- 130. Duffy, B. K., Dé-fago, G. 1999. Environmental factors modulating antibiotic and siderophore biosynthesis by *Pseudomonas fluorescens* biocontrol strains. *Appl Environ Microbiol* 65:2429–2438.
- 131. Strigul, N. S., Kravchenko, L. V. 2006. Mathematical modeling of PGPR inoculation into the rhizosphere. *Environ Model Soft* 21:1158–1171.
- 132. Pathma, J., Kamaraj, Kennedy, R., Sakthivel, N. 2011. Mechanisms of fluorescent pseudomonads that mediate biological control of phytopathogens and plant growth promotion of crop plants. In: Maheshwari DK (ed) Bacteria in agrobiology: plant growth responses. *Springer*, Berlin/ Heidelberg, pp 77–105.